GE Healthcare

illustra™ Ready-To-Go™ GenomiPhi™ V3 DNA Amplification Kit

Product booklet

Codes: 25-6601-24 25-6601-96 25-6601-97



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1.1. Product use restriction/warranty.

The **Ready-To-Go GenomiPhi V3 DNA Amplification Kit** components have been designed, developed, and sold **for research purposes only**. They are **suitable for** *in vitro* **use only**. Not recommended or intended for diagnosis of disease in humans or animals. No claim or representation is intended for its use to identify any specific organism or for clinical use (diagnostic, prognostic, therapeutic, or blood banking). Do not use internally or externally in humans or animals.

It is the responsibility of the user to verify the use of the **Ready-To-Go GenomiPhi V3 DNA Amplification Kit** for a specific application range as the performance characteristic of this kit has not been verified to a specific organism.

2. Handling, Safety Warnings and Precautions

2.1. Handling, Preparation, and Storage of Starting Materials

Warning: For research use only.

Not recommended or intended for diagnosis of disease in humans or animals. Do not use internally or externally in humans or animals.

This kit is sensitive to small amounts of DNA. Wear gloves and safety glasses to avoid contamination.

> All chemicals should be considered as potentially hazardous. We therefore recommend that this product is handled only by those persons who have been trained in laboratory techniques and that it is used in accordance with the principles of good laboratory practice. Wear suitable protective clothing

such as laboratory overalls, safety glasses and gloves. Care should be taken to avoid contact with skin or eyes. In the case of contact with skin or eyes wash immediately with water. See material safety data sheet(s) and/or safety statement(s) for specific advice.

Ready-To-Go GenomiPhi V3 DNA Amplification Kit is optimized for whole genome amplification from at least 10 ng of high quality genomic DNA template.

> Use of less DNA or low quality DNA (such as degraded DNA, or DNA from formalin fixed paraffin embedded samples) can result in amplification bias. Negligible amplification product is produced in the absence of template DNA up to 1.5 hours amplification.

Wear gloves at all times during the preparation to avoid contamination.

2.2. Storage Conditions

Store the kit at ambient temperature.

Once opened, completely reseal the pouch containing the Ready-To-Go GenomiPhi V3 DNA amplification kit cakes. To ensure maximum shelf life and optimal performance, storage in a desiccator is recommended

Open product may be stored by resealing the pouch and folding the sealed edge over several times and sealing with a clip.

Store reconstituted Control DNA (Lambda) at -20°C.

For longer-term stability, onceopened, store $2 \times$ denaturation buffer at 2–8°C.

2.3. Expiry

This product has been designed to deliver high quality results for up to 12 months from the date of manufacture. Please refer to the expiration date on the product label.

3. Components

3.1. Kit Contents

Ready-To-Go GenomiPhi V3 DNA Amplification Kit

| Cat. No. | 24 Reactions 25-6601-24 | 96 Reactions 25-6601-96 | 5 × 96 Reactions 25-6601-97 |
|---|----------------------------|----------------------------|--------------------------------|
| 2× Denaturation buffer | 1 × 1.2 ml | 1 x 1.2 ml | 5 x 1.2 ml |
| Ready-To-Go | 1 / 1.2 1111 | 1 \ 1.2 | 5 × 1.2 111 |
| GenomiPhi V3 | 3 × 8-well | 1 × 96-well | 5 × 96-well |
| cakes | strips | plate | plates |
| Control DNA (Lambda), 10ng/ul when reconstituted with 50 µl of sterile | | | |
| water | 1 × vial | 1 × vial | 1 × vial |
| Strips of 8 domed | | | |
| caps | 12 strips | 12 strips | 5 × 12 strips |

3.2. Reagents to be supplied by the user

- Liquid-handling supplies Sterile vials and pipette tips; pipettes, micro centrifuge
- Water Use PCR-grade water that is free of contaminating DNases or nucleic acid.
- Ice bucket or cold block for maintaining Ready-To-Go GenomiPhi V3 DNA Amplification Kit cakes at 4°C during reconstitution and prior to DNA amplification.
- Amplification reactions should be performed in the wells provided. Individual strips can be split from the rest of the plate by bending the strip upwards, **carefully** cutting the seal between adjacent columns of wells, then snapping-off the required strip of 8 wells.

For fewer reactions, carefully tap out cake into suitable PCR tube, taking precautions to avoid introducing contaminating DNA (as section 2.1)

 \bullet Thermocycler or water baths - for incubations at 30°C, 65°C and 95°C.

4. Product description

4.1. The Basic Principle

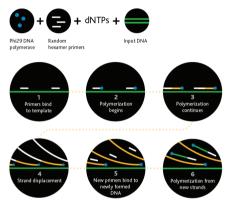


Figure 1. Overview of the Ready-To-Go GenomiPhi V3 DNA Amplification Kit procedure.

Figure 1 shows an overview of whole genome amplification by isothermal stand displacement using the **Ready-To-Go GenomiPhi V3 DNA Amplification Kit**. DNA is briefly heat-denatured in denaturation buffer then cooled. This is added to the freezedried cake which contains DNA polymerase, random hexamers, nucleotides, salts and buffers. Isothermal amplification proceeds at 30°C for 1.5 hours. After amplification the enzyme is heat inactivated during a 10 minute incubation at 65°C.

4.2. Kit specifications

Typical amplification kinetics with **Ready-To-Go GenomiPhi V3 DNA Amplification Kit** is shown in Figure 2. Microgram quantities of DNA are generated from nanogram amounts of starting material in 1.5 hours. Typical DNA yields from a **Ready-To-Go GenomiPhi V3** DNA Amplification Kit reaction are >12-20 µg per 20 µl reaction when starting with 10 ng of purified DNA. Kinetics will vary if crude or un-quantified samples are amplified. Increased reaction times (2 hours) may be helpful for samples such as crude blood or buccal swabs. Control reactions without added template DNA do not produce any product during 1.5 h reactions. The average product length is greater than 10 kb. DNA replication is extremely accurate due to the proofreading 3'-5' exonuclease activity of the DNA polymerase (3, 4).

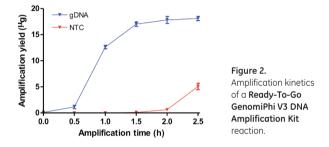


Figure 2 shows the comparison of amplification of 10 ng purified gDNA with no template control (NTC). Most commercial DNA isolation kits and homemade purification procedures produce suitable DNA for the amplification.

| Sample | See section | Typical DNA yield (µg) |
|---------------------------------------|-------------|---------------------------|
| 10 ng Lambda DNA | 5.2 | 18 |
| 10 ng Human genomic DNA - heat | 5.2 | 16 |
| 10 ng Human genomic DNA - chemical | 5.3 | 16 |
| Blood lysate | 5.4 | 15 |
| Dried blood spots | 5.5 | 18 |

Figure 3 table showing typical amplification yields for **Ready-To-Go GenomiPhi V3 DNA Amplification Kit** using either purified DNA (10 ng) or DNA amplified from blood or blood-spotted paper using protocols described later in the booklet.

5. Protocols

5.1. Short Protocol

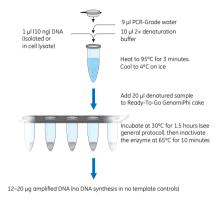


Figure 4. Schematic representation of Ready-To-Go GenomiPhi V3 DNA Amplification Kit protocol.

5.2. General Protocol

The following outlined steps describe a general protocol for amplifying template DNA.

This protocol should be considered a starting point for optimizing the reaction in your laboratory.

DNA should be resuspended in PCR-grade water or TE-1 buffer (10 mM Tris-HCl pH 8.0; 0.1 mM EDTA). The Denaturation buffer is supplied as a $2\times$ solution to allow some flexibility in volume of DNA that can be used.

Note: The total volume of template DNA and water should equal 10 $\mu l.$

When required, the Control DNA should be reconstituted with 50 μ l PCR-grade water. A screw-cap has been provided to re-seal the tube. Store at -20°C after reconstitution (in aliquots if so desired).

| | Mix 2× denaturation buffer with template DNA. Add 10 μl 2× Denaturation Buffer to 1 μl of 10 ng template DNA. | 10 µl denaturation buffer |
|---|--|------------------------------|
| | Add 9 µl PCR-grade water. | 9 µl PCR-grade water |
| ۲ | Template DNA should be resuspended in PCR-grade water or TE- ¹ buffer. If sample DNA concentration is less than 10 ng/µl more volume can be used, the total volume of DNA plus water should equal 10 µl. | 1 µl DNA template (10 ng) |
| | 2. Denature template DNA Heat the samples to 95°C for 3 minutes then cool to 4°C on ice. | 95°C for 3 minutes |
| | Heating the DNA for longer than 3 minutes or at higher temperatures can cause damage to the DNA. | |
| | 3. Reconstitute the Ready-To-Go GenomiPhi V3 cake with the denatured template DNA. Add the 20 μ l of denatured DNA template from Step 2 to each cake. Seal wells with domed caps provided. Keep each amplification reaction on ice prior to incubation at 30°C. | 20 µl denatured DNA |
| ۲ | The reconstituted Ready-To-Go GenomiPhi V3 cake contains all the components required for DNA amplification and will generate amplification products if exposed to temperatures > 4°C for sufficient time. | |
| | Incubate for DNA amplification. Incubate the samples at 30°C for 1.5 hours. | 30°C for 1.5 hours |

5. Inactivate the Phi29 DNA polymerase enzyme.

Heat the samples to 65°C for 10 minutes then 65°C for 10 minutes cool to 4°C

Heating is required to inactivate the exonuclease activity of the DNA polymerase which may otherwise begin to degrade the amplification product.

6. Storage of amplified material

Store amplification reactions at -20°C.

Ready-To-Go GenomiPhi DNA V3 amplification products should be stored and treated as genomic DNA. Minimize freeze-thaw cycles.

5.3. Chemical Denaturation and Amplification of Template DNA

The steps outlined below describe a general protocol for amplifying template DNA.

1. Prepare chemical denaturation solution. neutralization buffer and potassium chloride solution

Chemical Denaturation Solution 400 mM KOH 10 mM FDTA

Neutralization Buffer

400 mM HCl 600 mM Tris-HCl, pH 7.5 (prepared by mixing 4 ml of 1 M HCl and 6 ml of 1 M Tris-HCl pH 7.5)

Potassium Chloride

150 mM KCl

2. Denature template DNA.

Mix $1~\mu l$ of template (10 ng) with $1~\mu l$ of Chemical Denaturation Solution. Incubate at room temperature for 3 minutes

Mix by pipetting up and down. Do not vortex.

3. Neutralize DNA solution.

Add $1\,\mu l$ of Neutralization Buffer and store on ice.

Mix by pipetting up and down. Do not vortex.

4. Add PCR-grade water and potassium chloride

Add **7 μl** of PCR-grade water and **10 μl** 150 mM KCl.

Keep on ice.

5. Reconstitute the Ready-To-Go GenomiPhi V3 cake with the denatured template DNA.

Add the **20 µl** of denatured DNA template from **Step 4** to each cake. Seal wells with domed caps provided. Keep each amplification reaction on ice prior to incubation at 30°C.

The reconstituted Ready-To-Go GenomiPhi V3 cake contains all the components required for DNA amplification and will generate amplification products if exposed to temperatures > 4°C for sufficient time. 1 μl of DNA + 1 μl Denaturation solution RT for 3 minutes

1 μl of Neutralization buffer

7 μl of PCR-grade water + 10 μl of potassium chloride

20 µl denatured DNA

| 6. Incubate for DNA amplification. Incubate the samples at 30°C for 1.5 hours. | 30°C for 1.5 hours |
|--|---------------------|
| Inactivate the Phi29 DNA polymerase enzyme. Heat the samples to 65°C for 10 minutes then cool to 4°C. | 65°C for 10 minutes |
| Heating is required to inactivate the exonuclease activity of the DNA polymerase which would otherwise degrade the amplification product. | |
| 8. Storage of the amplified material. Store amplification reactions at -20°C. | |

Ready-To-Go GenomiPhi DNA V3 amplification products should be stored and treated as genomic DNA. Minimize freeze-thaw cycles.

5.4. DNA Amplification from Blood Cells

Note: Components of blood (e.g. heme) can be inhibitory to the GenomiPhi reaction.

Amplification is faster and more reproducible when the blood sample is diluted before amplification.

A reaction time of 2 hours is suggested.

1. Prepare cell lysis solution, neutralization buffer and potassium chloride solution.

Cell lysis solution 400 mM KOH 10 mM EDTA 100 mM DTT

Neutralization Buffer

400 mM HCl 600 mM Tris-HCl, pH 7.5 (prepared by mixing 4 ml of 1 M HCl and 6 ml of 1 M Tris-HCl pH 7.5)

Potassium Chloride

150 mM KCl

2. Cell lysis

Dilute blood 3 times with physiological buffer, e.g., PBS (137 mM NaCl; 2.7 mM KCl; 10 mM phosphate buffer pH 7.4).

Add one part of this mixture with one part of cell lysis solution.

Mix well with gentle tapping and incubate on ice for 10 minutes

Mix by pipetting up and down. Do not vortex.

3. Neutralize cell lysate.

Add one part of neutralization buffer to the cell lysate, mix well and store on ice.

buffer

Mix by pipetting up and down. Do not vortex.

4. Add PCR-grade water and potassium chloride.

To **3 µl** from **Step 3**, add **7 µl** of nuclease-free water and **10 µl** 150 mM KCl. Keep on ice.

5. Reconstitute the Ready-To-Go GenomiPhi V3 cake with the denatured template DNA.

Add the **20 µl** of denatured DNA template from

Dilute blood × 3

1 μl of diluted blood cells + 1 μl cell lysis solution.

10 minutes on ice

1 µl of Neutralization buffer

7 μl of PCR-grade water + 10 μl of potassium chloride

20 µl denatured DNA

| | Step 4 to each cake. Keep each amplification reaction on ice prior to incubation at 30°C. | |
|---|---|---------------------|
| ۲ | The reconstituted Ready-To-Go GenomiPhi V3 cake contains all the components required for DNA amplification and will generate amplification products if exposed to temperatures > 4°C for sufficient time. | |
| | 6. Incubate for DNA amplification. Incubate the samples at 30°C for 2.0 hours. | 30°C for 2.0 hours |
| | 7. Inactivate the Phi29 DNA polymerase enzyme. Heat the samples to 65°C for 10 minutes then | 65°C for 10 minutes |
| | cool to 4°C. | |
| ۲ | Heating is required to inactivate the exonuclease activity of the DNA polymerase which would otherwise degrade the amplification product. | |
| | Storage of the amplified material. Store amplification reactions at -20°C. | |
| | Ready-To-Go GenomiPhi DNA V3 amplification | |

Ready-To-Go GenomiPhi DNA V3 amplification products should be stored and treated as genomic DNA. Minimize freeze-thaw cycles.

5.5. DNA Amplification from Blood-blotted Paper (e.g. Whatman's FTA Paper or Guthrie card)

1. Prepare cell lysis solution, neutralization buffer, $2\times$ denaturation buffer and TE-¹ buffer.

Cell lysis / denaturation solution

400 mM KOH 10 mM FDTA 100 mM DTT

Neutralization Buffer

400 mM HCL 600 mM Tris-HCl, pH 7.5

2x Denaturation buffer

20 mM Hepes, pH8.25 10 mM FTDA 0.02% Tween-20 150 mM KCl

TE-¹ buffer 10 mM Tris-HCl pH 8.0

0.1 mM FDTA

2. Punch processing.

From the blood-blotted and dried FTA paper. punch out 1.2 or 3 mm diameter disks.

Wash the disks three times for 5 minutes with FTA purification reagent (GE catalog number WB120204) in a micro centrifuge tube (using 200 µl for 1.2 mm, and 500 µl for 3 mm disks).

Wash each disk twice for 5 minutes with TE-1 buffer. Remove as much fluid as possible at the end of the second TF-¹ buffer wash

Mix by pipetting up and down. Do not vortex.

3 Denaturation

Add 20 µl cell lysis / denaturation solution to 20 µl cell lysis solution the damp disks from Step 2.

Mix well with gentle tapping and incubate on ice for 10 minutes.

Mix by pipetting up and down. Do not vortex.

4. Neutralization.

Add 20 μl neutralization buffer to the cell lysate.

Add 20 μI PCR-grade water, mix well and store on ice.

Mix by pipetting up and down. Do not vortex.

5. Add 2× denaturation buffer.

To 10 μl from Step 4, add 10 μl 2× denaturation buffer.

Keep on ice.

6. Reconstitute the Ready-To-Go GenomiPhi V3 cake with the denatured cell lysate DNA.

Add the **20 µl** of denatured DNA template from ^{20 µl denatured DNA **Step 5** to each cake. Keep each amplification reaction on ice prior to incubation at 30°C.}

The reconstituted Ready-To-Go GenomiPhi V3 cake contains all the components required for DNA amplification and will

generate amplification products if exposed to temperatures > 4°C for sufficient time.

7. Incubate for DNA amplification. Incubate the samples at 30°C for 2 hours.

30°C for 2 hours

8. Inactivate the Phi29 DNA polymerase enzyme.

20 µl Neutralization Buffer + 20 µl PCR-grade water

65°C for 10 minutes

Heat the samples to 65°C for 10 minutes then cool to $4^{\circ}\text{C}.$

Heating is required to inactivate the exonuclease activity of the DNA polymerase which would otherwise degrade the amplification product.

9. Storage of the amplified material.

Store amplification reactions at -20°C.

Ready-To-Go GenomiPhi DNA V3 amplification products should be stored and treated as genomic DNA. Minimize freeze-thaw cycles.

5.6. Quantification of Amplification Products

Quantification is generally not required as every reaction will yield approximately the same amount of DNA. **Quant-iT™ PicoGreen® dsDNA quantification reagent** (Life Technologies, P7581) is recommended if accurate quantitation is required.

Quantification of non-purified amplification products by UV absorption will generate inaccurate results due to the presence of unused hexamers in the completed reaction.

1. Prepare TE buffer.

Dilute the concentrated 20 \times TE buffer included Prepare 1 \times TE buffer in the kit to 1 \times concentration using water.

▲ Use only sterile, DNase free water when preparing the dilution to ensure accurate quantification.

2. Prepare 1:25 dilution of PicoGreen reagent.

Determine the required volume of a 1:25 dilution of PicoGreen reagent.

- Volume = 100 µl/sample × # of samples Determine the volume of stock PicoGreen reagent necessary to produce the required volume of a 1:25 dilution
- Volume = $\frac{\text{volume of required dilution}}{25}$

Reagent adsorbs to glass surfaces. Use plastic ware only. Protect the solution from light at all times.

3. Prepare the λ DNA standard curve.

Dilute the λ DNA standard supplied in the Quant-iT PicoGreen kit to a **10 ng/µl** working solution. Use this working stock to prepare a standard curve (see table).

Add **100 µl** of each dilution to each well of the assay plate.

| Standard | λDNA | λDNA | 1 x TE |
|----------|------|------------|---------|
| Number | (ng) | (10 ng/µl) | |
| 1 | 600 | 60 µl | 40 µl |
| 2 | 500 | 50 µl | 50 µl |
| 3 | 400 | 40 µl | 60 µl |
| 4 | 200 | 20 µl | 80 µl |
| 5 | 100 | 10 µl | 90 µl |
| 6 | 50 | 5 µl | 95 µl |
| 7 | 25 | 2.5 µl | 97.5 µl |
| 8 | 0 | 0 µl | 100 µl |

4. Dilute the Ready-To-Go GenomiPhi V3 amplification products.

Dilute the **Ready-To-Go** GenomiPhi V3 amplification products 1:10 by **adding 180 μl** of 1 × TE to each amplification reaction.

Due to the viscosity of the amplification product, mix amplification products thoroughly by vortexing heavily.

5. Add diluted Ready-To-Go GenomiPhi amplification products to the assay plate .

Aliquot **95 \muI** of 1 × TE into each sample well.

Add 5 μl of diluted sample for a final volume of 100 $\mu l.$

Because the amplification product is diluted before the assay, the dilution factor must be taken into consideration when calculating total yields.

6. Add diluted PicoGreen to sample wells.

Add 100 µl. of the 1:25 dilution of PicoGreen to all wells containing standards and samples. Mix contents well by pipetting up and down.

Seal the plate with foil and spin in micro plate centrifuge for 1 minute to eliminate bubbles.

Protect plate from light at all times. The plate must be read 5–10 minutes after addition of PicoGreen reagent to ensure accurate quantification. 95 µl of TE + 5 µl of diluted sample

7. Measure the sample fluorescence.

Place the sample assay plate into a fluorescence micro plate reader.

Set the fluorescence reader at the following parameters:

- Excitation wavelength: 480 nm
- Emission wavelength: 520 nm
- Gain: Optimal

If it is not possible to set the instrument gain to optimal, find a way to have the instrument read the sample where the highest DNA concentration generates readings that fall within the linear dynamic range of the instrument.

8. Calculate the concentration of the amplification product.

Generate a standard curve of fluorescence versus DNA concentration. Determine the concentration of Ready-To-Go GenomiPhi V3 Kit amplified products from the equation of the line derived from the standard curve.

6. Appendix

6.1. Troubleshooting

| Problems | Possible cause and suggestions |
|--|---|
| Reduced yield/ no amplification product. | Contamination of template DNA Excessive contaminants carried over from the starting material can inhibit the DNA polymerase. Dilute or clean-up the DNA and re-amplify. Extending the amplification time will help when inhibitory material is causing reduced yields. |
| | Inactive Enzyme • It is critical that the Ready-To-Go GenomiPhi V3 DNA Amplification Kit freeze-dried cakes are stored properly. Once opened, completely reseal the pouch, fold the sealed edge over several times and seal with a clip. In high humidity environments, store unopened and resealed pouches in a desiccator to maximise product life. |
| | • Each cake should be a bright white solid having the shape of the base of the well. Cakes that have been exposed to moisture will become increasingly more translucent in appearance and this will impact both stability and performance. |
| | Perform a control (Lambda DNA) reaction to confirm performance of the Ready-To-Go GenomiPhi V3 DNA Amplification Kit. |

| Problems | Possible cause and suggestions |
|--|--|
| Reduced yield/ no amplification product. | Low quality DNA Amplification kinetics strongly favor intact templates. Avoid template preparation steps that can damage DNA. |
| | Prolonged denaturation Heating at 95°C for 3 minutes is sufficient to denature template DNA. Longer denaturation times can nick the template and decrease the amplification efficiency. |
| Poor performance in downstream applications. | Degraded/low amounts of template DNA In the absence of input DNA or poor quality of input DNA, there will be no or minimal DNA synthesis in the amplification reactions within 1.5 hours. |
| | Degraded or low amounts of starting DNA template may not amplify consistently or representatively. Increase the amount of starting DNA. |
| | Use high quality genomic DNA for amplification. |
| | Inhibition of optimized downstream conditions |
| | Components of the Ready-To-Go GenomiPhi V3 reaction may affect previously optimized conditions for some downstream applications. Purify the amplification products after amplification. |

6.2. References

- 1. Dean, F. et al., Genome Research 11, 1095–1099 (2001).
- 2. Lizardi, P. et al., Nat. Genet. 19, 225–232 (1998).
- 3. Estaban, J.A. et al., J. Biol. Chem. 268, 2719-2726 (1993).
- 4. Nelson, J.R. et al; BioTechniques 32, S44-S47 (2002).

6.3. Related Products

| GenomiPhi Products* | |
|--|--|
| illustra Ready-To-Go GenomiPhi HY (high yield) | 25-6603-24 25-6603-96 25-6603-97 |
| illustra GenomiPhi V2 (liquid format) | 25-6600-31 |
| illustra GenomiPhi HY (high yield, liquid format) | 25-6600-22 |
| DNA Purification Products* | |
| illustra tissue and cells genomicPrep Mini Spin Kit | 28-9042-76 |
| illustra tissue and cells genomicPrep Midi Flow Kit | 28-9042-73 |
| illustra blood genomicPrep Mini Spin Kit | 28-9042-64 |
| illustra blood genomicPrep Midi Flow Kit | 28-9042-61 |
| illustra triplePrep | 28-9425-44 |
| illustra bacteria genomicPrep Mini Spin Kit | 28-9042-58 |
| PCR Products* | |
| illustra PureTaq™ Ready-To-Go PCR Beads | 27-9559-01 |
| illustra Hot Start Mix Ready-To-Go | 28-9006-53 |
| illustra GFX™ PCR DNA and Gel Band Purification Kit | 28-9034-70 |
| illustra ExoProStar™ - PCR and Sequence Reaction Clean-Up | US78210 |

| illustra ExoProStar 1-Step - PCR and Sequence Reaction | 1 |
|--|------------|
| Clean-Up | US77702 |
| DNA Polymerase (cloned) | 27-0798-04 |
| illustra Solution dNTPs (multiple formats available) | 28-4065-52 |
| | |

* please see <u>http://www.gelifesciences.com</u> for an overview of available pack sizes.

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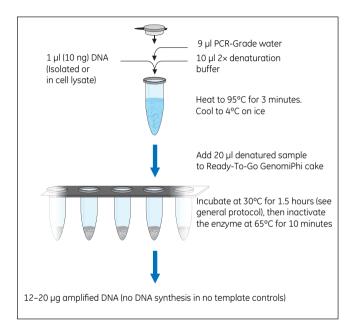
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Warning: For research use only. Not recommended or intended for diagnosis of disease in humans or animals. Do not use internally or externally in humans or animals.

1.1. Product use restriction/warranty.

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It is the responsibility of the user to verify the use of the **Ready-To-Go GenomiPhi V3 DNA Amplification Kit** for a specific application range as the performance characteristic of this kit has not been verified to a specific organism.

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